

2-ALKYLIDENE-18,19-DINOR-VITAMIN D COMPOUNDS

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2-ALKYLIDENE-18,19-DINOR-VITAMIN D COMPOUNDS

BACKGROUND OF THE INVENTION

[0001] This invention relates to vitamin D compounds, and more particularly to 2-alkylidene-18,19-dinor-vitamin D compounds, pharmaceutical uses for these compounds and a general method for chemically synthesizing these compounds.

[0002] The natural hormone, $1\alpha,25$ -dihydroxyvitamin D_3 and its analog in the ergosterol series, i.e. $1\alpha,25$ -dihydroxyvitamin D_2 are known to be highly potent regulators of calcium homeostasis in animals and humans, and their activity in cellular differentiation has also been established, Ostrem et al., Proc. Natl. Acad. Sci. USA, 84, 2610 (1987). Many structural analogs of these metabolites have been prepared and tested, including 1α -hydroxyvitamin D_3 , 1α -hydroxyvitamin D_2 , various side chain homologated vitamins and fluorinated analogs. Some of these compounds exhibit an interesting separation of activities in cell differentiation and calcium regulation. This difference in activity may be useful in the treatment of a variety of diseases as renal osteodystrophy, vitamin D-resistant rickets, osteoporosis, psoriasis, and certain malignancies.

[0003] A particularly interesting class of vitamin D analogs are referred to as the 19-nor-vitamin D compounds. The 19-nor-vitamin D compounds are characterized by the replacement of the A-ring exocyclic methylene group (carbon 19), typical of the vitamin D system, by two hydrogen atoms. Biological testing of such 19-nor-analogs (e.g., $1\alpha,25$ -dihydroxy-19-nor-vitamin D_3) revealed a selective activity profile with high potency in inducing cellular differentiation, and very low calcium mobilizing activity. Thus, these compounds are potentially useful as therapeutic agents for the treatment of malignancies, or the treatment of various skin disorders. Two different methods of synthesis of such 19-nor-vitamin D analogs have been described (Perlman et al., Tetrahedron Lett. 31, 1823 (1990);

Perlman et al., Tetrahedron Lett. 32, 7663 (1991), and DeLuca et al., U.S. Pat. No. 5,086,191).

[0004] In U.S. Pat. No. 4,666,634, 2 β -hydroxy and alkoxy (e.g., ED-71) analogs of 1 α ,25-dihydroxyvitamin D₃ have been described and examined by Chugai group as potential drugs for osteoporosis and as antitumor agents. See also Okano et al., Biochem. Biophys. Res. Commun. 163, 1444 (1989). Other 2-substituted (with hydroxyalkyl, e.g., ED-120, and fluoroalkyl groups) A-ring analogs of 1 α ,25-dihydroxyvitamin D₃ have also been prepared and tested (Miyamoto et al., Chem. Pharm. Bull. 41, 1111 (1993); Nishii et al., Osteoporosis Int. Suppl. 1, 190 (1993); Posner et al., J. Org. Chem. 59, 7855 (1994), and J. Org. Chem. 60, 4617 (1995)).

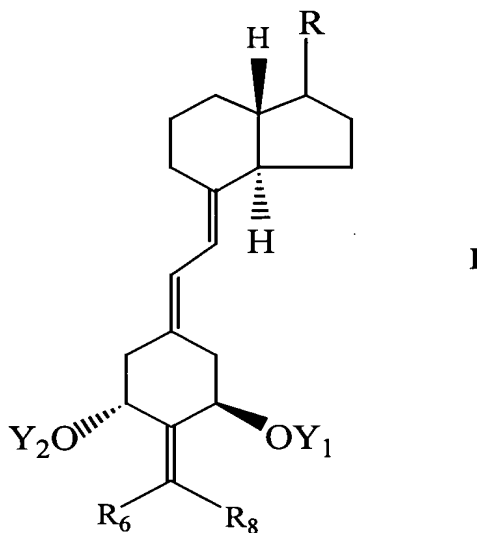
[0005] Recently, 2-substituted analogs of 1 α ,25-dihydroxy-19-nor-vitamin D₃ have also been synthesized, i.e. compounds substituted at the 2-position of the A-ring with hydroxy or alkoxy groups (DeLuca et al., U.S. Pat. No. 5,536,713), with 2-alkyl groups (DeLuca et al U.S. Patent No. 5,945,410), and with 2-alkylidene groups (DeLuca et al U.S. Patent No. 5,843,928), which exhibit interesting and selective activity profiles. All these studies indicate that binding sites in vitamin D receptors can accommodate different substituents at C-2 in the synthesized vitamin D analogs.

[0006] Another class of known vitamin D compounds are the 18,19-dinor analogs. These analogs have both the C-18 angular methyl substituent (carbon 18) normally attached to carbon 13 of the CD-ring structure and the C-19 exocyclic methylene group (carbon 19) normally attached to carbon 10 of the A-ring, which are typical of all vitamin D compounds, removed and replaced by hydrogen atoms. Reference should be made to the U.S. Patent 5,843,927 as well as U.S. Patents 5,756,489 and 5,721,225 for a more complete description of these compounds, their pharmaceutical uses, and their synthesis.

SUMMARY OF THE INVENTION

[0007] In a continuing effort to explore the 19-nor class of pharmacologically important vitamin D compounds, the present invention is directed toward 2-alkylidene-18,19-dinor-vitamin D analogs, various pharmaceutical uses for these compounds, and a general method for chemically synthesizing these compounds. In particular, the present invention is directed toward (20S)-2-methylene-1 α ,25-dihydroxy-18,19-dinor-vitamin D₃, its biological activity, and various pharmaceutical uses for this compound.

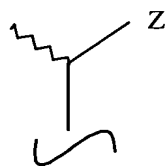
[0008] Structurally these novel 2-alkylidene-18,19-dinor-vitamin D analogs are characterized by the general formula I shown below:



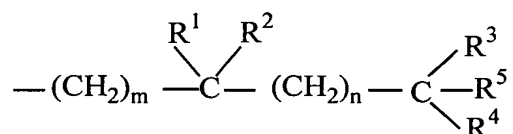
where Y₁ and Y₂, which may be the same or different, are each selected from the group consisting of hydrogen and a hydroxy-protecting group, R₆ and R₈, which may be the same or different, are each selected from the group consisting of hydrogen, alkyl, hydroxyalkyl and fluoroalkyl, or, when taken together represent the group $-(CH_2)_x-$ where X is an integer from 2 to 5, and where the group R represents any of the typical side chains known for vitamin D type compounds.

[0009] More specifically R can represent a saturated or unsaturated hydrocarbon radical of 1 to 35 carbons, that may be straight-chain, branched or cyclic and that may contain one or more additional substituents, such as hydroxy- or protected-hydroxy

groups, fluoro, carbonyl, ester, epoxy, amino or other heteroatomic groups. Preferred side chains of this type are represented by the structure below



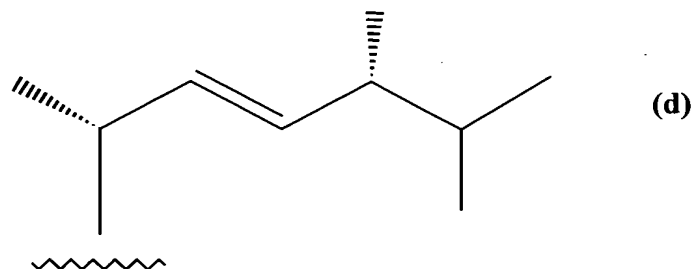
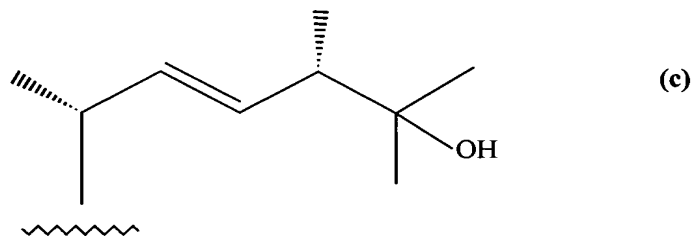
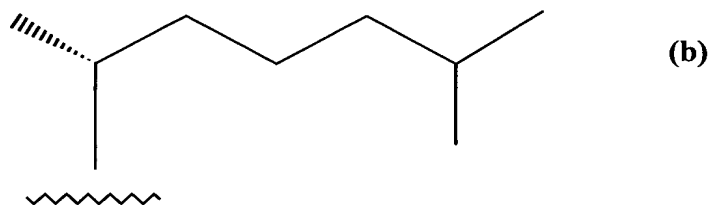
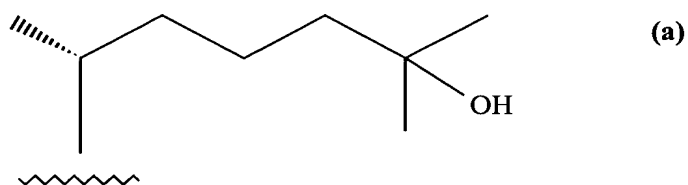
where the stereochemical center (corresponding to C-20 in steroid numbering) may have the R or S configuration, (i.e. either the natural configuration about carbon 20 or the 20-epi configuration), and where Z is selected from Y, -OY, -CH₂OY, -C≡CY and -CH=CHY, where the double bond may have the cis or trans geometry, and where Y is selected from hydrogen, methyl, -COR⁵ and a radical of the structure:

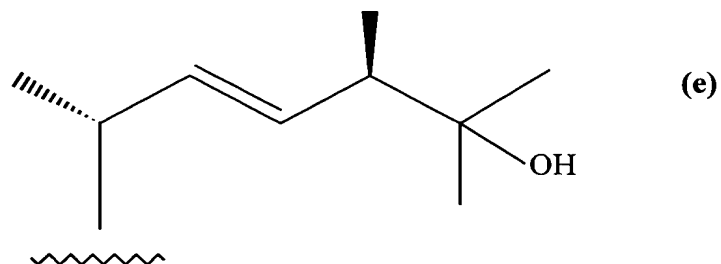


where m and n, independently, represent the integers from 0 to 5, where R¹ is selected from hydrogen, deuterium, hydroxy, protected hydroxy, fluoro, trifluoromethyl, and C₁₋₅-alkyl, which may be straight chain or branched and, optionally, bear a hydroxy or protected-hydroxy substituent, and where each of R², R³, and R⁴, independently, is selected from deuterium, deuteroalkyl, hydrogen, fluoro, trifluoromethyl and C₁₋₅ alkyl, which may be straight-chain or branched, and optionally, bear a hydroxy or protected-hydroxy substituent, and where R¹ and R², taken together, represent an oxo group, or an alkylidene group, =CR²R³, or the group - (CH₂)_p-, where p is an integer from 2 to 5, and where R³ and R⁴, taken together, represent an oxo group, or the group -(CH₂)_q-, where q is an integer from 2 to 5, and where R⁵ represents hydrogen, hydroxy, protected hydroxy, or C₁₋₅ alkyl and wherein any of the CH-groups at positions 20, 22, or 23 in the side chain may be replaced by a nitrogen atom, or where any of the groups -CH(CH₃)-, -(CH₂)_m-, (CH₂)_n or -CR¹R²- at positions 20, 22, and 23, respectively, may be replaced by an oxygen or sulfur atom.

[0010] The wavy line to the methyl substituent at C-20 indicates that carbon 20 may have either the R or S configuration, i.e. the natural configuration (20R) or the unnatural 20-epi configuration (20S).

[0011] Specific important examples of side chains with natural 20*R*-configuration are the structures represented by formulae (a), b), (c), (d) and (e) below. i.e. the side chain as it occurs in 25-hydroxyvitamin D₃ (a); vitamin D₃ (b); 25-hydroxyvitamin D₂ (c); vitamin D₂ (d); and the C-24 epimer of 25-hydroxyvitamin D₂ (e):





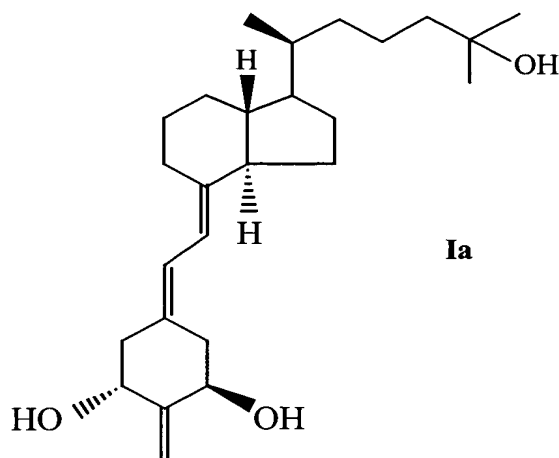
[0012] The above novel 2-alkylidene-18,19-dinor vitamin D compounds exhibit a desired, and highly advantageous, pattern of biological activity. These compounds are characterized by relatively high intestinal calcium transport activity, i.e. similar to that of $1\alpha,25$ -dihydroxyvitamin D_3 , while also exhibiting relatively low activity, as compared to $1\alpha,25$ -dihydroxyvitamin D_3 , in their ability to mobilize calcium from bone. Hence, these compounds are highly specific in their calcemic activity. Their preferential activity on intestinal calcium transport and reduced calcium mobilizing activity allows the in vivo administration of these compounds for the treatment and prophylaxis of metabolic bone diseases where bone loss is a major concern. Because of their preferential calcemic activity on gut calcium transport, these compounds would be preferred therapeutic agents for the treatment and prophylaxis of diseases where bone formation is desired, such as osteoporosis, especially low bone turnover osteoporosis, steroid induced osteoporosis, senile osteoporosis or postmenopausal osteoporosis, as well as osteomalacia and renal osteodystrophy. The compounds may be administered transdermally, orally or parenterally. The compounds may be present in a pharmaceutical composition in an amount from about $0.01\mu\text{g/gm}$ to about $100\mu\text{g/gm}$ of the composition, preferably from about $0.1\mu\text{g/gm}$ to about $50\mu\text{g/gm}$ of the composition, and may be administered in dosages of from about $0.01\mu\text{g/day}$ to about $100\mu\text{g/day}$, preferably from about $0.1\mu\text{g/day}$ to about $50\mu\text{g/day}$.

[0013] The compounds of the invention are also especially suited for treatment and prophylaxis of human disorders which are characterized by an imbalance in the immune system, e.g. in autoimmune diseases, including multiple sclerosis, diabetes mellitus, lupus, host versus graft reaction, and rejection of transplants; and additionally

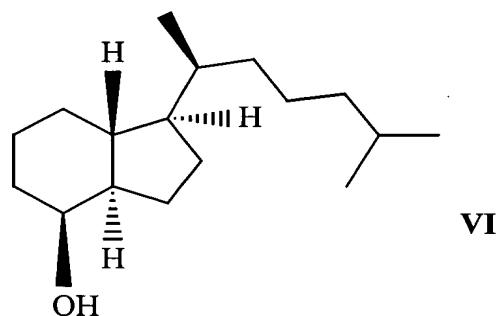
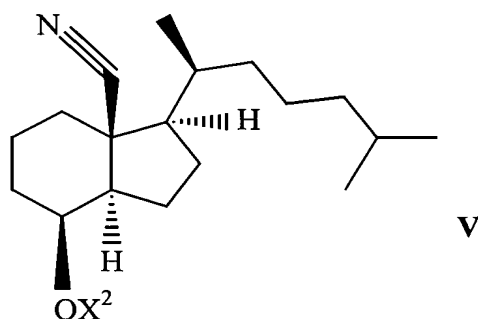
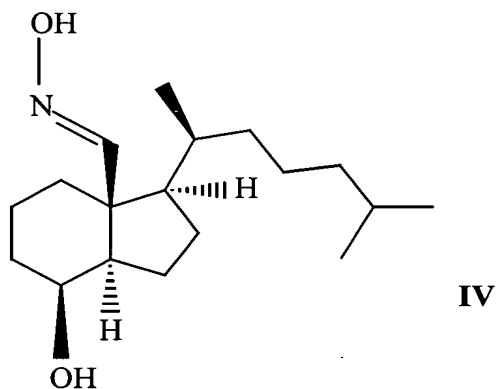
for the treatment and prophylaxis of inflammatory diseases, such as rheumatoid arthritis, asthma, and inflammatory bowel diseases such as Crohn's disease or ulcerative colitis, as well as the improvement of bone fracture healing and improved bone grafts. It has also been discovered that these compounds increase breaking strength (cortical strength) as well as crushing strength (trabecular strength) of bones. Thus, these compounds could also be used in conjunction with bone replacement procedures such as hip replacements, knee replacements, and the like. Acne, alopecia, skin conditions such as dry skin (lack of dermal hydration), undue skin slackness (insufficient skin firmness), insufficient sebum secretion and wrinkles, and hypertension are other conditions which may be treated with the compounds of the invention.

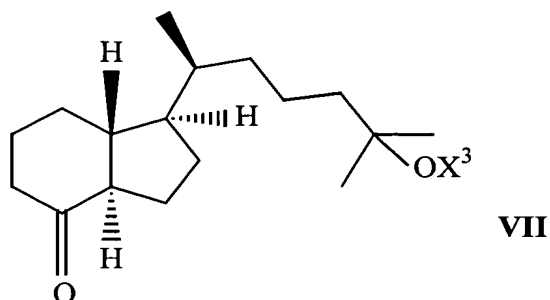
[0014] The above compounds are also characterized by high cell differentiation activity. Thus, these compounds also provide therapeutic agents for the treatment of psoriasis, or as an anti-cancer agent, especially against leukemia, colon cancer, breast cancer, skin cancer and prostate cancer. The compounds may be present in a composition to treat psoriasis in an amount from about 0.01 μ g/gm to about 100 μ g/gm of the composition, preferably from about 0.1 μ g/gm to about 50 μ g/gm of the composition, and may be administered topically, transdermally, orally or parenterally in dosages of from about 0.01 μ g/day to about 100 μ g/day, preferably from about 0.1 μ g/day to about 50 μ g/day.

[0015] In particular, 2-methylene-18,19-dinor-(20S)-1 α ,25-dihydroxy-vitamin D₃ has been synthesized and its binding, transcriptional, calcemic (both intestinal calcium transport and bone calcium mobilization) and differentiation activities determined. Structurally this 18,19-dinor analog is characterized by the general formula Ia shown below:



[0016] The invention also provides a novel synthesis for the production of the end products of formula I, and specifically of formula Ia. In addition, this invention provides novel intermediate compounds formed during the synthesis of the end products. Structurally, these novel intermediates are characterized by the general formulae IV, V, VI and VII, below where X^1 may be $-H$ or $-NO$, and X^2 and X^3 may be $-H$ or a hydroxy protecting group.





BRIEF DESCRIPTION OF THE DRAWINGS

[0017] Figure 1 is a graph illustrating the relative activity of $1\alpha,25$ -dihydroxyvitamin D_3 (C001) as well as the herein described and claimed (20S)-2-methylene-18,19-dinor- $1\alpha,25$ -dihydroxyvitamin D_3 (DP035) in binding to the $1\alpha,25$ -dihydroxyvitamin D pig intestinal nuclear receptor;

[0018] Figure 2 is a graph illustrating the percent HL-60 cell differentiation as a function of the concentration of $1\alpha,25$ -dihydroxyvitamin D_3 (C001), (20S)-2-methylene-19-nor- $1\alpha,25$ -dihydroxyvitamin D_3 (2MD) and (20S)-2-methylene-18,19-dinor- $1\alpha,25$ -dihydroxyvitamin D_3 (DP035);

[0019] Figure 3 is a graph illustrating the transcriptional activity as a function of the concentration of $1\alpha,25$ -dihydroxyvitamin (C001), (20S)-2-methylene-19-nor- $1\alpha,25$ -dihydroxyvitamin D_3 (2MD) and (20S)-2-methylene-18,19-dinor- $1\alpha,25$ -dihydroxyvitamin D_3 (DP035);

[0020] Figure 4 is a bar graph illustrating the intestinal calcium transport activity of (20S)-2-methylene-18,19-dinor- $1\alpha,25$ -dihydroxyvitamin D_3 (DP035) at various dosages as compared to control (vehicle) and $1\alpha,25$ -dihydroxyvitamin D_3 (C001);

[0021] Figure 5 is a bar graph illustrating the bone calcium mobilization activity of (20S)-2-methylene-18,19-dinor- $1\alpha,25$ -dihydroxyvitamin D_3 (DP035) at various dosages as compared to control (vehicle) and $1\alpha,25$ -dihydroxyvitamin D_3 (C001); and

[0022] Figure 6 is a bar graph illustrating the bone calcium mobilization activity of (20S)-2-methylene-18,19-dinor-1 α ,25-dihydroxyvitamin D₃ (DP035) as compared to 1 α ,25-dihydroxyvitamin D₃ (C001) at various dosages.

DETAILED DESCRIPTION OF THE INVENTION

[0023] As used in the description and in the claims, the term "hydroxy-protecting group" signifies any group commonly used for the temporary protection of hydroxy functions, such as for example, alkoxycarbonyl, acyl, alkylsilyl or alkylarylsilyl groups (hereinafter referred to simply as "silyl" groups), and alkoxyalkyl groups. Alkoxycarbonyl protecting groups are alkyl-O-CO- groupings such as methoxycarbonyl, ethoxycarbonyl, propoxycarbonyl, isopropoxycarbonyl, butoxycarbonyl, isobutoxycarbonyl, tert-butoxycarbonyl, benzyloxycarbonyl or allyloxycarbonyl. The term "acyl" signifies an alkanoyl group of 1 to 6 carbons, in all of its isomeric forms, or a carboxyalkanoyl group of 1 to 6 carbons, such as an oxalyl, malonyl, succinyl, glutaryl group, or an aromatic acyl group such as benzoyl, or a halo, nitro or alkyl substituted benzoyl group. The word "alkyl" as used in the description or the claims, denotes a straight-chain or branched alkyl radical of 1 to 10 carbons, in all its isomeric forms. Alkoxyalkyl protecting groups are groupings such as methoxymethyl, ethoxymethyl, methoxyethoxymethyl, or tetrahydrofuranyl and tetrahydropyranyl. Preferred silyl-protecting groups are trimethylsilyl, triethylsilyl, t-butyl dimethylsilyl, dibutylmethylsilyl, diphenylmethylsilyl, phenyl dimethylsilyl, diphenyl-t-butylsilyl and analogous alkylated silyl radicals. The term "aryl" specifies a phenyl-, or an alkyl-, nitro- or halo-substituted phenyl group.

[0024] A "protected hydroxy" group is a hydroxy group derivatised or protected by any of the above groups commonly used for the temporary or permanent protection of hydroxy functions, e.g. the silyl, alkoxyalkyl, acyl or alkoxycarbonyl groups, as previously defined. The terms "hydroxyalkyl", "deuteroalkyl" and "fluoroalkyl" refer

to an alkyl radical substituted by one or more hydroxy, deuterium or fluoro groups respectively.

[0025] It should be noted in this description that the term "24-homo" refers to the addition of one methylene group and the term "24-dihomo" refers to the addition of two methylene groups at the carbon 24 position in the side chain. Likewise, the term "trihomo" refers to the addition of three methylene groups. Also, the term "26,27-dimethyl" refers to the addition of a methyl group at the carbon 26 and 27 positions so that for example R^3 and R^4 are ethyl groups. Likewise, the term "26,27-diethyl" refers to the addition of an ethyl group at the 26 and 27 positions so that R^3 and R^4 are propyl groups.

[0026] In the following lists of side chain unsaturated and side chain saturated compounds, if the methyl group attached at the carbon 20 position is in its epi or unnatural configuration, the term "20(S)" or "20-epi" should be included in each of the following named compounds. Also, if the side chain contains an oxygen atom substituted at any of positions 20, 22 or 23, the term "20-oxa," "22-oxa" or "23-oxa," respectively, should be added to the named compound. The named compounds could also be of the vitamin D₂ type if desired.

[0027] Specific and preferred examples of the 2-alkylidene-18,19-dinor-vitamin D compounds of structure I when the side chain is unsaturated are:

[0028] 2-methylene-18,19-dinor-1 α -hydroxy-22-dehydrovitamin D₃;

[0029] 2-methylene-18,19-dinor-25-hydroxy-22-dehydrovitamin D₃;

[0030] 2-methylene-18,19-dinor-1 α ,25-dihydroxy-22-dehydrovitamin D₃;

[0031] 2-methylene-18,19-dinor-24-homo-1,25-dihydroxy-22-dehydrovitamin D₃;

[0032] 2-methylene-18,19-dinor-24-dihomo-1,25-dihydroxy-22-dehydrovitamin D₃;

- [0033]** 2-methylene-18,19-dinor-24-trihomo-1,25-dihydroxy-22-dehydrovitamin D₃;
- [0034]** 2-methylene-18,19-dinor-26,27-dimethyl-24-homo-1,25-dihydroxy-22-dehydrovitamin D₃;
- [0035]** 2-methylene-18,19-dinor-26,27-dimethyl-24-dihomo-1,25-dihydroxy-22-dehydrovitamin D₃;
- [0036]** 2-methylene-18,19-dinor-26,27-dimethyl-24-trihomo-1,25-dihydroxy-22-dehydrovitamin D₃;
- [0037]** 2-methylene-18,19-dinor-26,27-diethyl-24-homo-1,25-dihydroxy-22-dehydrovitamin D₃;
- [0038]** 2-methylene-18,19-dinor-26,27-diethyl-24-dihomo-1,25-dihydroxy-22-dehydrovitamin D₃;
- [0039]** 2-methylene-18,19-dinor-26,27-diethyl-24-trihomo-1,25-dihydroxy-22-dehydrovitamin D₃;
- [0040]** 2-methylene-18,19-dinor-26,27-dipropyl-24-homo-1,25-dihydroxy-22-dehydrovitamin D₃;
- [0041]** 2-methylene-18,19-dinor-26,27-dipropyl-24-dihomo-1,25-dihydroxy-22-dehydrovitamin D₃; and
- [0042]** 2-methylene-18,19-dinor-26,27-dipropyl-24-trihomo-1,25-dihydroxy-22-dehydrovitamin D₃.
- [0043]** With respect to the above unsaturated compounds, it should be noted that the double bond located between the 22 and 23 carbon atoms in the side chain may be in either the (E) or (Z) configuration. Accordingly, depending upon the configuration, the term "22,23(E)" or "22,23(Z)" could be included in each of the above named compounds. Also, it is common to designate the double bond located between the 22 and 23 carbon atoms with the designation " Δ^{22} ". Thus, for example, the fourth named compound above could also be written as 2-methylene-18,19-dinor-24-homo-22,23(E)-

Δ^{22} -1,25-(OH)₂D₃ where the double bond is the (E) configuration. Similarly, if the methyl group attached at carbon 20 is in the unnatural configuration, this compound could be written as 2-methylene-18,19-dinor-20(S)-24-homo-22,23(E)- Δ^{22} -1,25-(OH)₂D₃.

[0044] Specific and preferred examples of the 2-alkylidene-18,19-dinor-vitamin D compounds of structure I when the side chain is saturated are:

[0045] 2-methylene-18,19-dinor-1 α -hydroxyvitamin D₃;

[0046] 2-methylene-18,19-dinor-25-hydroxyvitamin D₃;

[0047] 2-methylene-18,19-dinor-1 α ,25-dihydroxyvitamin D₃;

[0048] 2-methylene-18,19-dinor-24-homo-1,25-dihydroxyvitamin D₃;

[0049] 2-methylene-18,19-dinor-24-dihomo-1,25-dihydroxyvitamin D₃;

[0050] 2-methylene-18,19-dinor-24-trihomo-1,25-dihydroxyvitamin D₃;

[0051] 2-methylene-18,19-dinor-26,27-dimethyl-24-homo-1,25-dihydroxyvitamin D₃;

[0052] 2-methylene-18,19-dinor-26,27-dimethyl-24-dihomo-1,25-dihydroxyvitamin D₃;

[0053] 2-methylene-18,19-dinor-26,27-dimethyl-24-trihomo-1,25-dihydroxyvitamin D₃;

[0054] 2-methylene-18,19-dinor-26,27-diethyl-24-homo-1,25-dihydroxyvitamin D₃;

[0055] 2-methylene-18,19-dinor-26,27-diethyl-24-dihomo-1,25-dihydroxyvitamin D₃;

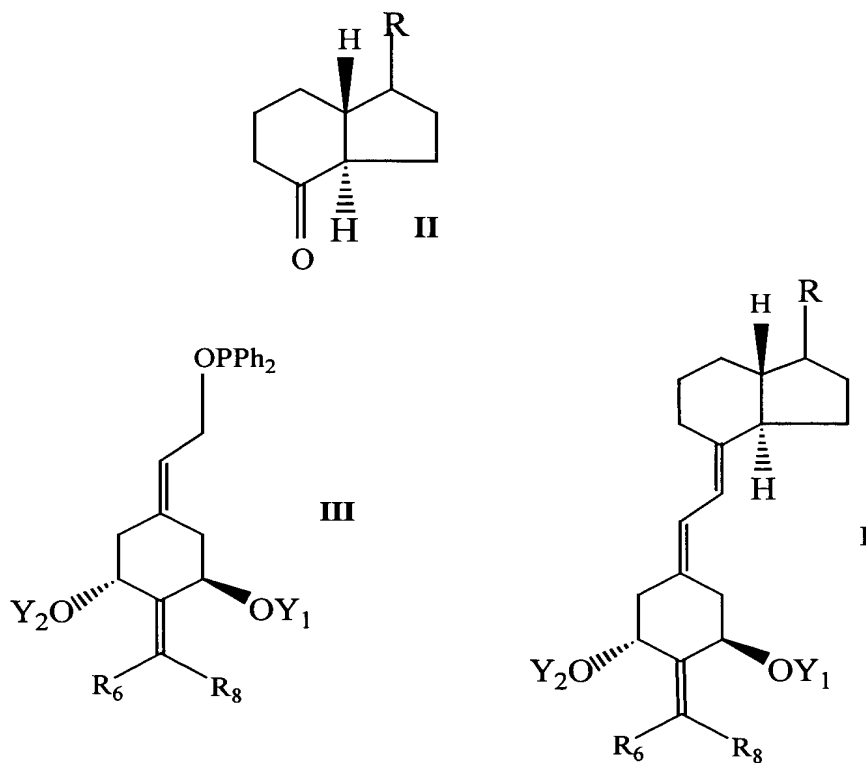
[0056] 2-methylene-18,19-dinor-26,27-diethyl-24-trihomo-1,25-dihydroxyvitamin D₃;

[0057] 2-methylene-18,19-dinor-26,27-dipropyl-24-homo-1,25-dihydroxyvitamin D₃;

[0058] 2-methylene-18,19-dinor-26,27-dipropyl-24-dihomo-1,25-dihydroxyvitamin D₃; and

[0059] 2-methylene-18,19-dinor-26,27-dipropyl-24-trihomo-1,25-dihydroxyvitamin D₃.

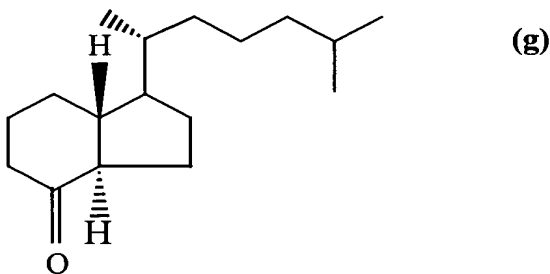
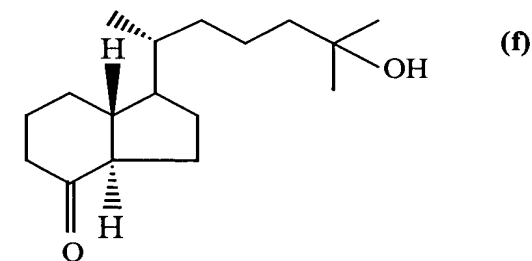
[0060] The preparation of 2-alkylidene-18,19-dinor vitamin D compounds having the structure I is based on the Wittig-Horner reaction of an 18-nor-CD-ring ketone (see (a) Baggiolini et al, J. Org. Chem., 1986, 51, 3098-3108; (b) Baggiolini et al, J. Am. Chem. Soc., 1982, 104, 2945-2948; and (c) Cohen et al, J. Org. Chem., 1979, 44, 3077-3080) and a phosphine oxide, i.e. the condensation of a bicyclic 18-nor-CD-ring type ketone II with an allylic phosphine oxide III to the corresponding 2-alkylidene-18,19-dinor vitamin D analog I followed by deprotection at C-1 and C-3 in the latter compounds:

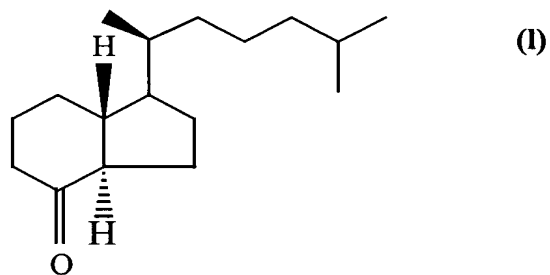
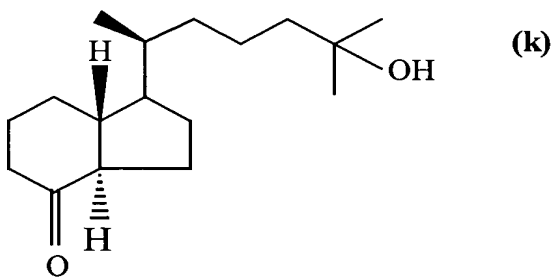
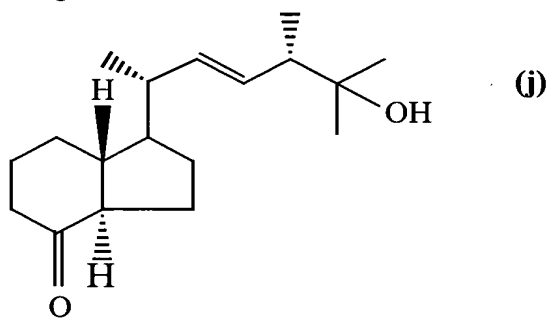
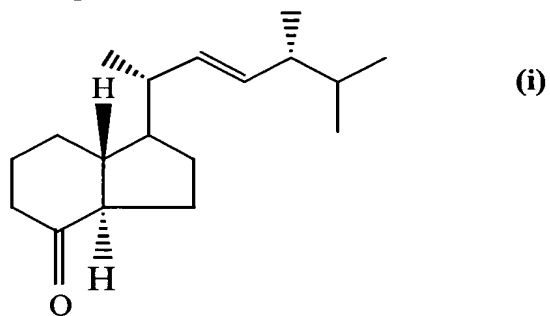
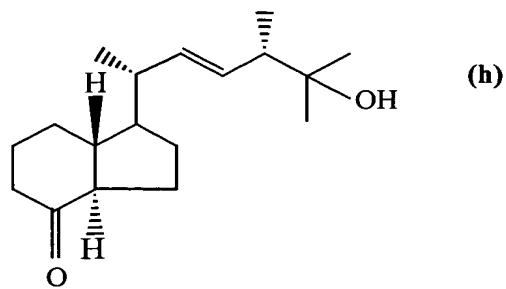


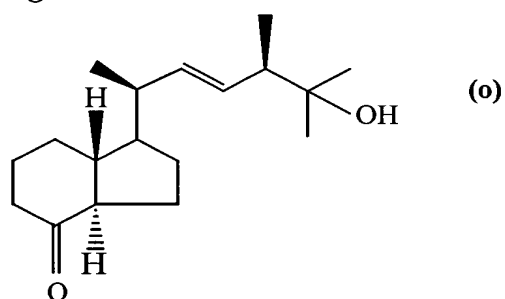
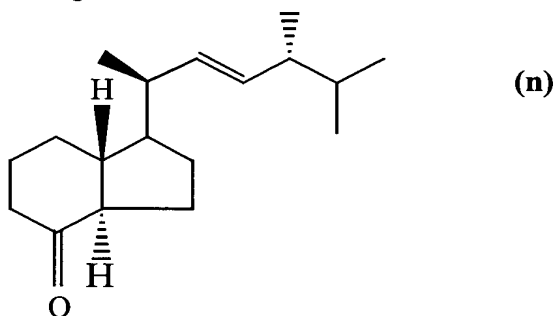
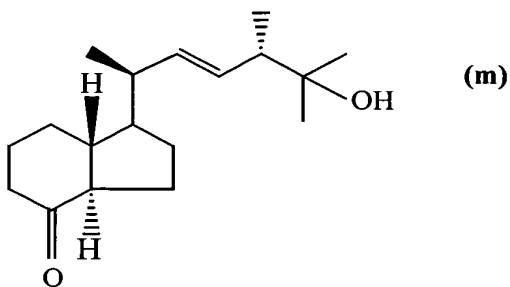
[0061] In the structures I, II and III groups R₆ and R₈, Y₁ and Y₂, and R represent groups defined above; Y₁ and Y₂ are preferably hydroxy-protecting groups

such as tert-butyldimethylsilyl (TBDMS), it being also understood that any functionalities in R that might be sensitive, or that interfere with the condensation reaction, be suitably protected as is well-known in the art. The process shown above represents an application of the convergent synthesis concept, which has been applied effectively for the preparation of vitamin D compounds [e.g. Lythgoe et al., J. Chem. Soc. Perkin Trans. I, 590 (1978); Lythgoe, Chem. Soc. Rev. 9, 449 (1983); Toh et al., J. Org. Chem. 48, 1414 (1983); Baggiolini et al., J. Org. Chem. 51, 3098 (1986); Sardina et al., J. Org. Chem. 51, 1264 (1986); J. Org. Chem. 51, 1269 (1986); DeLuca et al., U.S. Pat. No. 5,086,191; DeLuca et al., U.S. Pat. No. 5,536,713].

[0062] Hydrindanones of the general structure II can be prepared starting from vitamin D₂ by the method of SCHEME 1 disclosed hereinafter. Specific important examples of such bicyclic 18-nor-CD ketones are the structures with the side chains (a), (b), (c), (d) and (e) described above, i.e. 25-hydroxy ketone (f); ketone (g); 25-hydroxy ketone (h); ketone (i) and 24-epi ketone (j). Other important 18-nor CD ketones of general structure II are the structures with the side chains (f) through (j) wherein the 20-methyl group is in its unnatural 20-epi configuration, i.e. ketones (k) through (o).







[0063] For the preparation of the required phosphine oxides of general structure III, a synthetic route has been developed starting from a diol, easily obtained from commercial (1R,3R,4S,5R)-(-)-quinic acid as described by Sicinski et al, J. Med. Chem., 1998, 41, 4462-4674. The overall process of transformation of the starting diol into the desired 2-alkylidene-A-ring synthon of general structure III, and more particularly, the 2-methylene-A-ring synthon 15 shown in SCHEME 1, is summarized and illustrated in U.S. Patent 6,843,928, the description of which is specifically incorporated herein by reference. Thus, the starting diol will be oxidized with ruthenium tetroxide to the corresponding hydroxyketone. The latter compound will be treated with an ylide prepared from methyltriphenylphosphonium bromide and n-butyllithium. The product of the Wittig reaction will be reduced by lithium aluminum hydride to a vicinal diol, which will be cleaved by sodium periodate, and the resulting

ketone will be converted to an unsaturated ester by the Peterson olefination with methyl (trimethylsilyl)acetate. The ester will then be reduced with DIBALH to an allylic alcohol which will be *in situ* tosylated with *n*-butyllithium and *p*-toluenesulfonyl chloride, converted into the corresponding phosphine by a reaction with diphenylphosphine lithium salt, and oxidized with hydrogen peroxide to the desired A-ring phosphine oxide 15. The Wittig-Horner coupling of the two fragments 14 and 15, to give the protected vitamin compound 16, followed by the deprotection of hydroxy groups in any known manner such as with tetrabutylammonium fluoride, will give the final analog 17.

[0064] Numerous 2-alkylidene-18,19-dinor-vitamin D compounds of the general structure I may be synthesized using the A-ring synthon III and the appropriate 18-nor-CD-ring ketone II having the desired side chain structure R. Thus, for example, Wittig-Horner coupling of the A-ring phosphine oxide 15 with *n*-butyllithium and any of the ketones (f), (g), (h), (i), (j), (k), (l), (m), (n) and (o) previously illustrated herein (or any other ketone with the desired side chain defined by R) may be performed as illustrated in SCHEME 1 to give the respective protected vitamin compound. This, after deprotection then affords the desired 2-methylene-18,19-dinor-vitamin D analog having the desired side chain structure R.

[0065] The C-20 epimerization may be accomplished by the analogous coupling of the phosphine oxide of structure III with the appropriate protected (20*S*)-CD-ring ketone of structure II which after hydrolysis of the hydroxy-protecting groups will give the desired (20*S*)-2-alkylidene-18,19-dinor-vitamin D analog having the desired side chain structure R.

[0066] As noted above, other 2-alkylidene-18,19-dinor-vitamin D analogs may be synthesized by the method disclosed herein. For example, 1 α -hydroxy-2-methylene-18,19-dinor-vitamin D₃ can be obtained by providing the CD-ring ketone (g).

[0067] This invention is described by the following illustrative examples. In these examples specific products identified by Arabic numerals (e.g. 1, 2, 3, etc) refer to the specific structures so identified in the preceding description and in SCHEME 1.

EXAMPLE 1

[0068] Preparation of (20S)-2-methylene-1 α ,25-dihydroxy-18,19-dinor-vitamin D₃ (17) via condensation (SCHEME I).

[0069] **Des-*A,B*-23,24-dinorcholane-8 β ,22-diol (1).** A solution of vitamin D₂ (5 g, 12.7 mmol) in methanol (400 mL) and pyridine (5 mL) was cooled to -78°C while purging with argon. The argon stream was stopped and a stream of ozone was passed until a blue color appeared. The solution was purged with oxygen until blue color disappeared and treated with NaBH₄ (1.2 g, 32 mmol). After 20 min. the second portion of NaBH₄ (1.2 g, 32 mmol) was added and reaction was allowed to warm to room temperature. The third portion of NaBH₄ (1.2 g, 32 mmol) was added and the reaction mixture was stirred overnight at room temperature. The reaction was quenched with 70 mL of water and concentrated under vacuum. The residue was extracted with methylene chloride (3 x 100 mL). The organic phase was washed with 1M aqueous solution of HCl (2 x 100 mL), saturated aqueous solution of NaHCO₃ (100 mL), dried over anhydrous MgSO₄ and concentrated under vacuum. The residue was purified by flash chromatography (25% ethyl acetate/hexane) to yield 1.875 g (8.84 mmol, 70% yield) of diol **1** as white crystals. $[\alpha]_D + 56.0$ (c 0.95, CHCl₃); mp 110-111°C; ¹H NMR (400 MHz, CDCl₃) δ 0.96 (3H, s), 1.03 (3H, d, $J = 6.6$ Hz), 3.38 (1H, dd, $J = 10.5$ Hz, $J = 6.8$ Hz), 3.64 (1H, dd, $J = 10.5$ Hz, $J = 3.2$ Hz), 4.09 (1H, d, $J = 2.3$ Hz); ¹³C NMR (100 MHz, CDCl₃) δ 13.6, 16.6, 17.4, 22.6, 26.6, 33.5, 38.2, 40.2, 41.3, 52.3, 52.9, 67.8, 69.2; MS (EI) m/z 212 (2, M⁺), 194 (17), 179 (18), 163 (10), 135 (19), 125 (34), 111 (100); exact mass calculated for C₁₃H₂₂O ([M - H₂O]⁺) 194.1671, found 194.1665.

[0070] **Des-*A,B*-8 β -(benzoyloxy)-23,24-dinorcholane-22-ol (2).** Diol **1** (1.85 g, 8.79 mmol) was dissolved in pyridine (30 mL) and DMAP (45 mg, 0.3 mmol) was added. The solution was cooled to 0°C then benzoyl chloride (3 mL, 3.6 g, 25 mmol) was added dropwise. The reaction mixture was kept at 5°C for 24 h. Methylene chloride (100 mL) was added and the resulting mixture was washed with 5% aqueous solution of HCl (100 mL), saturated aqueous solution of CuSO₄ (2 x 80 mL), saturated aqueous solution of NaHCO₃ (80 mL) and water (100 mL). The extract was dried over anhydrous MgSO₄. Removal of the solvent in vacuo afforded a crude dibenzoate.

[0071] The crude dibenzoate (5.05 g) was added at room temperature to a solution of KOH (87%, 1.5 g, 23.3 mmol) in absolute ethanol (30 mL). The resulting reaction mixture was stirred at room temperature for 3 h 20 min. Then the reaction mixture was quenched with ice and neutralized with 5% aqueous solution of HCl. The reaction mixture was extracted with methylene chloride (3 x 60 mL). The combined organic phases were washed with saturated aqueous solution of NaHCO₃ (50 mL) and dried over anhydrous MgSO₄. Drying agent was removed and solvent was evaporated in vacuo. Pure product was obtained by column chromatography (25% ethyl acetate/hexane) to give 2.58 g (8.16 mmol, 93% yield from diol **1**) of monobenzoate **2**. [α]_D + 65.2 (c 1.15, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 3.39 (1H, dd, J = 10.4 Hz, J = 6.8 Hz), 3.65 (1H, dd, J = 10.5 Hz, J = 3.2 Hz), 5.42 (1H, br d, J = 22.2 Hz), 7.45 (2H, m), 7.56 (1H, m), 8.05 (2H, m); ¹³C NMR (100 MHz, CDCl₃) δ 13.6, 16.6, 18.0, 22.7, 26.6, 30.5, 38.4, 39.8, 41.9, 51.4, 52.7, 67.7, 72.1, 128.3, 129.5, 130.8, 166.5; MS (EI) m/z 211 (4), 194 (52), 179 (11), 135 (41), 108 (23), 105 (100); exact mass (ESI) calculated for C₂₀H₂₈O₃Na ([M + Na]⁺) 339.1936, found 339.1941.

[0072] **Des-*A,B*-8 β -(benzoyloxy)-23,24-dinorcholane-22-al (3).** Sulfur trioxide pyridine complex (7.02 g, 44.1 mmol) was added to a solution of alcohol **2**

(2.32 g, 7.34 mmol) and triethylamine (5.15 mL, 3.71 g, 36.7 mmol) in anhydrous methylene chloride (30 mL) and DMSO (8 mL) at 0°C. The reaction mixture was stirred under argon for 20 min. at 0°C and then concentrated in vacuo. The residue was purified by column chromatography (5% ethyl acetate/hexane) to give 2.05 g (6.53 mmol, 90% yield) of aldehyde **3**. $[\alpha]_D + 67.4$ (c 0.95, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.10 (3H, s), 1.15 (3H, d, *J* = 6.8 Hz), 5.44 (1H, br d, *J* = 2.2 Hz), 7.45 (2H, m), 7.56 (1H, m), 8.05 (2H, m), 9.60 (1H, d, *J* = 3.2 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 13.6, 14.1, 18.1, 23.1, 26.2, 30.7, 39.8, 42.6, 49.2, 51.2, 51.5, 128.6, 129.7, 130.9, 133.0, 205.0; MS (EI) *m/z* 285 (3), 216 (3), 208 (9), 180 (17), 162 (47), 147 (21), 135 (46), 122 (16), 105 (100), 95 (22), 77 (49); exact mass (ESI) calculated for C₁₉H₂₅O₂ ([M – CHO]⁺) 285.1855, found 285.1848.

[0073] (20*R*)-Des-*A,B*-8β-(benzoyloxy)-23,24-dinorcholane-22-ol (4**).** To a solution of aldehyde **3** (2.05 g, 6.53 mmol) in methylene dichloride (25 mL), 40% aqueous solution of *n*-Bu₄NOH (8.4 mL, 12.9 mmol) was added. The resulting reaction mixture was vigorously stirred overnight. Methylene dichloride (30 mL) was then added and the mixture was washed with water (20 mL), dried over anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by column chromatography (5% ethyl acetate/hexane) to give 1.50 g (4.78 mmol) of the mixture of diastereoisomeric aldehydes.

[0074] The mixture of aldehydes was dissolved in ethanol (15 mL) and NaBH₄ (350 mg, 9.2 mmol) was added. The resulting mixture was stirred for 30 min. The reaction mixture was quenched with saturated aqueous solution of NH₄Cl (30 mL). The mixture was extracted with methylene dichloride (3 x 40 mL) and the combined organic phases were washed with water (30 mL), dried over anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by column chromatography (5% ethyl acetate/hexane) to give 870 mg (2.75 mmol, 42% yield) of **4** and 437 mg (1.38 mmol, 21% yield) of **2**. $[\alpha]_D + 50.0$ (c 1.10,

CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.97 (3H, d, *J* = 6.7 Hz), 1.07 (3H, s), 3.48 (1H, dd, *J* = 10.5 Hz, *J* = 7.1 Hz), 3.76 (1H, dd, *J* = 10.6 Hz, *J* = 3.5 Hz), 5.42 (1H, s), 7.45 (2H, m), 7.55 (1H, m), 8.05 (2H, m); ¹³C NMR (125 MHz, CDCl₃) δ 13.9, 16.5, 18.0, 22.5, 26.4, 30.5, 37.5, 39.3, 41.7, 51.5, 52.7, 66.9, 72.0, 128.3, 129.5, 130.8, 166.5; MS (EI) *m/z* 316 (16, M⁺), 301 (5), 285 (9), 242 (11), 194 (60), 147 (71), 105 (100); exact mass (ESI) calculated for C₂₀H₂₈O₃Na ([M + Na]⁺) 339.1936, found 339.1948.

[0075] (20*R*)-Des-*A,B*-8β-(benzoyloxy)-23,24-dinor-22-(tosyloxy)cholane (5). To a mixture of alcohol 4 (870 mg, 2.75 mmol), triethylamine (1.5 mL, 10.8 mmol) and DMAP (20 mg) in anhydrous methylene dichloride (20 mL) tosyl chloride (710 mg, 3.73 mmol) was added at 0°C. The reaction mixture was allowed to stand at room temperature for 16 h. Then methylene dichloride (100 mL) was added and the mixture was washed with saturated aqueous solution of NaHCO₃ (2 x 50 mL), dried over anhydrous MgSO₄ and concentrated under reduced pressure. The residue was purified by column chromatography (5% ethyl acetate/hexane) to give 1162 mg (2.47 mmol, 90% yield) of 5. [α]_D + 14.2 (c 0.95, CHCl₃); mp. 100-102°C; ¹H NMR (500 MHz, CDCl₃) δ 0.90 (3H, d, *J* = 6.6 Hz), 0.98 (3H, s), 2.46 (3H, s), 3.83 (1H, dd, *J* = 9.2 Hz, *J* = 7.2 Hz), 4.15 (1H, dd, *J* = 9.3 Hz, *J* = 3.3 Hz), 7.35 (2H, d, *J* = 8.1 Hz), 7.44 (2H, m), 7.55 (1H, m), 7.80 (2H, d, *J* = 8.1 Hz), 8.02 (2H, m); ¹³C NMR (125 MHz, CDCl₃) δ 13.9, 16.6, 17.9, 21.6, 22.3, 26.3, 30.4, 34.8, 39.1, 41.6, 71.8, 74.0, 127.9, 128.4, 129.5, 129.7, 130.7, 132.8, 133.1, 144.6, 166.7; MS (EI) *m/z* 365 (12), 348 (61), 193 (9), 176 (32), 161 (13), 134 (19), 105 (100), 91 (17), 77 (20); exact mass (ESI) calculated for C₂₇H₃₄O₅SNa ([M + Na]⁺) 493.2025, found 493.2032..

[0076] (20*S*)-Des-*A,B*-cholestan-8β-ol (7). Magnesium turnings (4.41 g, 184 mmol) were stirred with a magnetic stir bar overnight under argon. Anhydrous THF (50 mL) and 1-chloro-3-methylbutane (11 mL, 90.8 mmol) were then added.

The mixture was refluxed for 6 h. The resulting solution of Grignard reagent **6** was then added via cannula to a stirred solution of **5** in anhydrous THF (15 mL) at -78°C followed by addition of a solution of dilithium tetrachlorocuprate (620 mg, 2.73 mmol) in anhydrous THF (27 mL). The cooling bath was removed and the reaction mixture was stirred overnight. The reaction mixture was poured into a stirred mixture of ice (15 mL) and saturated aqueous solution of NH₄Cl (40 mL). The mixture was then extracted with ethyl acetate (3 x 100 mL), washed with water and dried over anhydrous Na₂SO₄. The residue was purified by column chromatography (5 to 25% ethyl acetate/hexane) to give 389 mg (1.46 mmol, 58% yield) of **7**. [α]_D + 9.6 (c 1.15, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.82 (3H, d, *J* = 6.6 Hz), 0.87 (6H, d, *J* = 6.6 Hz), 0.93 (3H, s), 4.07 (1H, s); ¹³C NMR (125 MHz, CDCl₃) δ 13.8, 17.5, 18.5, 22.4, 22.5, 22.6, 22.7, 24.0, 27.1, 28.0, 29.7, 33.6, 34.8, 35.5, 39.4, 40.3, 41.9, 52.7, 56.3, 69.5; MS (EI) *m/z* 266 (45, M⁺), 251 (19), 233 (8), 177 (9), 163 (11), 152 (20), 135 (30), 125 (37), 111 (100); exact mass calculated for C₁₈H₃₄O 266.26310, found 266.2623.

[0077] (20S)-Des-A,B-cholestan-8 β -yl nitrite (8). A solution of **7** (185 mg, 0.69 mmol) in chloroform (5 mL) was treated with tert-butyl nitrite (1 mL) for 1 h in darkness. Benzene (10 mL) was then added and solvents were removed under reduced pressure, protecting the mixture from light. ¹H NMR (500 MHz, CDCl₃) δ 0.76 (3H, s), 0.81 (3H, d, *J* = 6.5 Hz), 0.87 (6H, d, *J* = 6.6 Hz), 5.78 (1H, s); ¹³C NMR (125 MHz, CDCl₃) δ 13.1, 17.9, 18.5, 22.2, 22.6, 22.7, 23.9, 27.1, 28.0, 31.5, 34.9, 35.3, 39.3, 39.7, 41.9, 51.9, 56.0.

[0078] (18E)-(20S)-18-(Hydroxyimino)-des-A,B-cholestan-8 β -ol (9). Crude nitrite was dissolved in anhydrous benzene (150 mL) and irradiated in an apparatus consisting of a Pyrex vessel with a watercooled immersion well and Hanovia high-pressure mercury arc lamp equipped with Pyrex filter. A slow stream of argon was passed through solution and temperature was maintained at about

10°C. A reaction progress was monitored by TLC. After 30 min. reaction was completed. Benzene was removed under reduced pressure and the residue was dissolved in 2-propanol (5 mL) and refluxed for 2 h, cooled and allowed to stand overnight to accomplish isomerisation of a nitroso compound to an oxime. The solvent was evaporated and the residue was purified on Waters silica gel Sep-Pack cartridge (25% ethyl acetate/hexane) to give 102 mg (0.35 mmol, 51% yield from 7) of the oxime **9**. $[\alpha]_D + 8.2$ (c 0.80, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 0.84 (3H, d, *J* = 6.3 Hz), 0.87 (6H, d, *J* = 6.6 Hz), 2.20 (1H, br d, *J* = 13.1 Hz), 4.04 (1H, br d, *J* = 2.6 Hz), 7.33 (1H, s), 10.8 (1H, br s); ¹³C NMR (100 MHz, CDCl₃) δ 17.5, 18.6, 21.8, 22.6, 22.7, 24.1, 27.2, 28.0, 34.3, 35.0, 35.6, 39.3, 49.5, 52.6, 56.7, 67.6, 152.2; MS (EI) *m/z* 295 (2, M⁺), 278 (28), 260 (20), 245 (8), 206 (19), 183 (38), 165 (13), 148 (15), 121 (100); exact mass calculated for C₁₈H₃₃NO₂Na ([M + Na]⁺) 318.2409, found 318.2412.

[0079] (20S)-8β-(Acetoxy)-des-A,B-cholestan-18-nitrile (10). A solution of **9** (100 mg, 0.34 mmol) in acetic anhydride (5 mL) was refluxed for 1.5 h. The reaction mixture was cooled, poured carefully into ice and extracted with benzene (3 x 40 mL). The combined organic phases were washed with saturated aqueous solution of NaHCO₃ (2 x 40 mL), water (30 mL), dried over anhydrous Na₂SO₄ and evaporated. The residue was purified on a Waters silica gel Sep-Pack cartridge (5% ethyl acetate/hexane) to give 91 mg (0.28 mmol, 84% yield) of **9**. $[\alpha]_D - 26.4$ (c 0.75, CHCl₃); IR (CHCl₃) 2228, 1741, 1241; ¹H NMR (500 MHz, CDCl₃) δ 0.87 (6H, d, *J* = 6.6 Hz), 0.91 (3H, d, *J* = 6.6 Hz), 2.15 (3H, s), 2.46 (1H, br d, *J* = 3.2 Hz), 5.20 (1H, s); ¹³C NMR (125 MHz, CDCl₃) δ 17.9, 18.8, 22.6, 22.7, 23.3, 23.8, 27.1, 28.0, 29.9, 35.6, 36.2, 36.3, 39.1, 45.6, 51.9, 54.1, 68.7, 121.2, 171.0; MS (EI) *m/z* 319 (18, M⁺), 304 (10), 290 (3), 277 (84), 259 (100), 244 (54), 234 (27), 216 (40), 202 (33), 188 (60), 174 (47), 147 (39), 134 (34), 121 (95); exact mass (ESI) calculated for C₂₀H₃₃NO₂Na ([M + Na]⁺) 342.2409, found 342.2413.

[0080] (20S)-Des-A,B-cholestan-18-nitrile-8 β -ol (11). **10** (90 mg, 0.28 mmol) was dissolved in methanol (3 mL) and treated with 5% solution of MeONa in methanol (3mL) for 2 h. The reaction mixture was quenched with a saturated aqueous solution of NH₄Cl (5 mL), water (10 mL), extracted with methylene dichloride (5 x 40 mL), dried over anhydrous Na₂SO₄ and evaporated. The residue was purified on a Waters silica gel Sep-Pack cartridge (20% ethyl acetate/hexane) to give 73 mg (0.26 mmol, 94% yield) of **10**. [α]_D – 6.1 (c 0.75, CHCl₃); IR (CHCl₃) 3486, 2228; ¹H NMR (500 MHz, CDCl₃) δ 0.87 (6H, d, *J* = 6.6 Hz), 0.92 (3H, d, *J* = 6.7 Hz), 2.43 (1H, br d, *J* = 3.1 Hz), 4.10 (1H, s); ¹³C NMR (125 MHz, CDCl₃) δ 17.9, 22.6, 22.7, 22.9, 23.9, 27.1, 28.0, 32.8, 35.7, 36.2, 36.3, 44.7, 53.4, 54.2, 122.5; MS (EI) *m/z* 277 (28, M⁺), 262 (34), 259 (18), 248 (16), 244 (24), 220 (30), 216 (18), 206 (100); exact mass calculated for C₁₈H₃₁NO 277.2496, found 277.2395.

[0081] (20S)-Des-A,B-18-norcholestan-8 β -ol (12). To a stirred mixture of potassium (110 mg, 2.82 mmol) in HMPA (280 μ l, 1.62 mmol) and diethyl ether (700 μ l) a solution of **11** (70 mg, 0.25 mmol) in *tert*-butyl alcohol (65 μ l) and diethyl ether (250 μ l) was added dropwise at 0°C under argon. The mixture was allowed to warm up to room temperature and stirred for 5 h. Remaining potassium was removed, a few drops of 2-propanol and benzene (20 mL) were added. Organic phase was washed with water (10 mL), dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified on Waters silica gel Sep-Pack cartridge (10% ethyl acetate/hexane) to give 54 mg (0.21 mmol, 85% yield) of **12**. [α]_D + 32.6 (c 0.90, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.78 (3H, d, *J* = 6.8 Hz), 0.87 (6H, d, *J* = 6.6 Hz), 4.06 (1H, s); ¹³C NMR (125 MHz, CDCl₃) δ 14.7, 20.2, 22.7, 22.9, 24.7, 25.3, 28.0, 30.8, 33.1, 33.5, 36.3, 39.3, 39.7, 48.6, 50.3, 67.9; MS (EI) *m/z* 252 (6, M⁺), 234 (21), 219 (23), 209 (26), 191 (8), 179 (4),

167 (13), 149 (89), 139 (47), 122 (90), 107 (35), 95 (80), 79 (87), 67 (88), 58 (100); exact mass calculated for $C_{17}H_{32}O$ 252.2453, found 252.2448.

[0082] (20S)-Des-*A,B*-25-hydroxy-18-norcholestane-8-one (13). To a stirred solution of $RuCl_3 \times H_2O$ (10 mg, 0.05 mmol) and $NaIO_4$ (227 mg, 1.06 mmol) in water (1 mL) a solution of **12** (74 mg, 0.29 mmol) in tetrachloromethane (0.75 mL) and acetonitrile (0.75 mL) was added. The reaction mixture was vigorously stirred for 3 days. Then a few drops of 2-propanol and water (10 mL) were added. Reaction products were extracted with methylene dichloride (3 x 20 mL). Organic phase was dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The residue was purified on Waters silica gel Sep-Pack cartridge (10 to 30% ethyl acetate/hexane) to give 13 mg (0.05 mmol, 17% yield) of **13**. 1H NMR (400 MHz, $CDCl_3$) δ 0.78 (3H, d, J = 6.7 Hz), 1.22 (6H, s), 2.01 (1H, br d, J = 12.3 Hz); ^{13}C NMR (100 MHz, $CDCl_3$) δ 14.3, 21.3, 22.2, 22.6, 27.8, 29.3, 29.7, 33.0, 36.5, 41.6, 44.1, 49.6, 51.0, 58.0, 71.0, 212.0; MS (EI) m/z 264 (3), 248 (57), 233 (19), 215 (4), 208 (15), 163 (29), 137 (100); exact mass (ESI) calculated for $C_{17}H_{30}O_2Na$ ($[M + Na]^+$) 289.2144, found 289.2136.

[0083] (20S)-25-[(Triethylsilyl)oxy]-des-*A,B*-18-norcholestane-8-one (14). To a stirred solution of **13** (12 mg, 45 μ mol) and 2,6-lutidine (13 μ L, 100 μ mol) in anhydrous methylene dichloride (250 μ L) triethylsilyl trifluoromethanesulfonate was added dropwise at -50°C under argon. After 20 min. a few drops of wet methylene dichloride and water (7 mL) were added. Reaction mixture was extracted with methylene dichloride (3 x 7 mL). Organic phase was dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The residue was purified on Waters silica gel Sep-Pack cartridge (3% ethyl acetate/hexane) and on HPLC (5% ethyl acetate/hexane, 4 mL/min., Zorbax-silica 10 x 250mm) to give 13 mg (34 μ mol, 76% yield) of **14**. 1H NMR (500 MHz, $CDCl_3$) δ 0.56 (6H, q, J = 7.9 Hz), 0.77 (3H, d, J = 6.8 Hz), 0.94 (9H, t, J = 7.9 Hz), 1.19 (6H, s); ^{13}C NMR (125

MHz, CDCl₃) δ 6.8, 7.1, 14.3, 21.4, 22.2, 22.7, 27.8, 29.7, 29.8, 29.9, 32.9, 36.4, 41.6, 45.2, 49.6, 51.1, 58.0, 73.4, 212.1; MS (EI) m/z 365 (8), 351 (100), 322 (6), 239 (2), 231 (25), 220 (4), 205 (15), 189 (4), 173 (92); exact mass (ESI) calculated for C₂₃H₄₄O₂SiNa ([M + Na]⁺) 403.3008, found 403.2995.

[0084] (20*S*)-2-Methylene-1 α ,25-dihydroxy-18,19-dinorvitamin D₃ (17).

To a stirred solution of phosphine oxide **15** (46 mg, 79 μ mol) in anhydrous THF (600 μ l) a 1.5 M solution of phenyl lithium in THF (63 μ l, 95 μ mol) was added at -20°C under argon. The mixture was stirred for 20 min. and then cooled to -78°C. A precooled solution of **14** (13 mg, 34 μ mol) in anhydrous THF (300 μ l) was added via cannula and the reaction mixture was stirred for 3 h at -78°C. After that the reaction mixture was stirred at 4°C overnight. Then ethyl acetate was added and organic phase was washed with brine, dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified on Waters silica gel Sep-Pack cartridge (hexane to 2% ethyl acetate/hexane) and then on HPLC (0.05% 2-propanol/hexane, 4 mL/min., Zorbax-silica 10 x 250mm) to give 13.5 mg (18 μ mol, 53% yield) of protected vitamin D₃ **16**. UV (hexane) λ_{max} = 242, 251, 261 nm; ¹H NMR (500 MHz, CDCl₃) δ 0.06 (3H, s), 0.11 (3H, s), 0.17 (3H, s), 0.19 (3H, s), 0.56 (6H, q, J = 8.0 Hz), 0.76 (3H, d, J = 6.7 Hz), 0.94 (9H, t, J = 8.0 Hz), 2.18 (1H, dd, J = 12.5 Hz, J = 8.1 Hz), 2.86 (1H, br d, J = 13.8 Hz), 4.42 (2H, m), 4.93 (1H, s), 4.96 (1H, s), 5.92 (1H, d, J = 11.1 Hz), 6.19 (1H, d, J = 11.1 Hz); ¹³C NMR (125 MHz, CDCl₃) δ -5.1, -4.9, -4.9, -4.8, 6.8, 7.1, 18.2, 18.2, 22.3, 23.1, 25.8, 25.8, 27.8, 29.0, 29.7, 29.8, 29.9, 31.3, 33.6, 36.5, 38.7, 45.3, 47.5, 49.0, 50.2, 52.3, 71.9, 72.3, 73.4, 106.3, 113.7, 122.4, 132.9, 143.8, 152.9; MS (EI) m/z 687 (6), 628 (2), 612 (100), 583 (6), 555 (4), 480 (29), 366 (44); exact mass calculated for C₄₀H₇₅O₃Si₃ ([M - t-Bu]⁺) 687.5024, found 687.5028.

[0085] **16** (13 mg, 17 μ mol) was dissolved in anhydrous THF (5 mL). Then a 1 M solution of tetrabutyl ammonium fluoride in THF (260 μ l, 260 μ mol) was added

dropwise followed by addition of activated molecular sieves 4A (200 mg). The reaction mixture was stirred under argon for 2 h. Then solvent was removed under reduced pressure and the residue was purified on Waters silica gel Sep-Pack cartridge (40 to 50% ethyl acetate/hexane). Crude **17** was then purified on HPLC (20% 2-propanol/hexane, 4 mL/min., Zorbax-silica 10 x 250mm) to give 3.8 mg (9.5 μ mol, 56% yield) of **17** at R_t = 5.58 min.; UV (EtOH) λ_{max} = 242, 250, 260 nm; ^1H NMR (500 MHz, CDCl_3) δ 0.77 (3H, d, J = 6.6 Hz), 1.21 (6H, s), 2.58 (1H, dd, J = 13.2 Hz, J = 3.9 Hz), 2.81 (1H, dd, J = 13.3 Hz, J = 4.4 Hz), 2.87 (1H, br d, J = 13.9 Hz), 4.48 (2H, m), 5.10 (1H, s), 5.11 (1H, s), 5.97 (1H, d, J = 11.3 Hz), 6.35 (1H, d, J = 11.3 Hz); MS (EI) m/z 402 (39, M^+), 384 (41), 366 (14), 351 (11), 299 (58), 231 (36), 142 (58), 69 (100); exact mass calculated for $\text{C}_{26}\text{H}_{42}\text{O}_3$ 402.3134, found 402.3121.

[0086] For treatment purposes, the novel compounds of this invention defined by formula I may be formulated for pharmaceutical applications as a solution in innocuous solvents, or as an emulsion, suspension or dispersion in suitable solvents or carriers, or as pills, tablets or capsules, together with solid carriers, according to conventional methods known in the art. Any such formulations may also contain other pharmaceutically-acceptable and non-toxic excipients such as stabilizers, anti-oxidants, binders, coloring agents or emulsifying or taste-modifying agents.

[0087] The compounds may be administered orally, topically, parenterally or transdermally. The compounds are advantageously administered by injection or by intravenous infusion or suitable sterile solutions, or in the form of liquid or solid doses via the alimentary canal, or in the form of creams, ointments, patches, or similar vehicles suitable for transdermal applications. Doses of from 0.01 μ g to 100 μ g per day of the compounds, preferably from about 0.1 μ g/day to about 50 μ g/day, are appropriate for treatment purposes, such doses being adjusted according to the disease to be treated, its severity and the response of the subject as is well understood in the art. Since the new compounds exhibit specificity of

action, each may be suitably administered alone, or together with graded doses of another active vitamin D compound -- e.g. 1α -hydroxyvitamin D₂ or D₃, or $1\alpha,25$ -dihydroxyvitamin D₃ -- in situations where different degrees of bone mineral mobilization and calcium transport stimulation is found to be advantageous.

[0088] Compositions for use in the above-mentioned treatment of psoriasis and other malignancies comprise an effective amount of one or more 2-alkylidene-18,19-dinor-vitamin D compound as defined by the above formula I as the active ingredient, and a suitable carrier. An effective amount of such compounds for use in accordance with this invention is from about 0.01 μ g to about 100 μ g per gm of composition, preferably from about 0.1 μ g/gm to about 50 μ g/gm of the composition, and may be administered topically, transdermally, orally or parenterally in dosages of from about 0.01 μ g/day to about 100 μ g/day, preferably from about 0.1 μ g/day to about 50 μ g/day.

[0089] The compounds may be formulated as creams, lotions, ointments, topical patches, pills, capsules or tablets, or in liquid form as solutions, emulsions, dispersions, or suspensions in pharmaceutically innocuous and acceptable solvent or oils, and such preparations may contain in addition other pharmaceutically innocuous or beneficial components, such as stabilizers, antioxidants, emulsifiers, coloring agents, binders or taste-modifying agents.

[0090] The compounds are advantageously administered in amounts sufficient to effect the differentiation of promyelocytes to normal macrophages. Dosages as described above are suitable, it being understood that the amounts given are to be adjusted in accordance with the severity of the disease, and the condition and response of the subject as is well understood in the art.

[0091] The formulations of the present invention comprise an active ingredient in association with a pharmaceutically acceptable carrier therefore and optionally other therapeutic ingredients. The carrier must be "acceptable" in the sense of being

compatible with the other ingredients of the formulations and not deleterious to the recipient thereof.

[0092] Formulations of the present invention suitable for oral administration may be in the form of discrete units as capsules, sachets, tablets or lozenges, each containing a predetermined amount of the active ingredient; in the form of a powder or granules; in the form of a solution or a suspension in an aqueous liquid or non-aqueous liquid; or in the form of an oil-in-water emulsion or a water-in-oil emulsion.

[0093] Formulations for rectal administration may be in the form of a suppository incorporating the active ingredient and carrier such as cocoa butter, or in the form of an enema.

[0094] Formulations suitable for parenteral administration conveniently comprise a sterile oily or aqueous preparation of the active ingredient which is preferably isotonic with the blood of the recipient.

[0095] Formulations suitable for topical administration include liquid or semi-liquid preparations such as liniments, lotions, applicants, oil-in-water or water-in-oil emulsions such as creams, ointments or pastes; or solutions or suspensions such as drops; or as sprays.

[0096] For asthma treatment, inhalation of powder, self-propelling or spray formulations, dispensed with a spray can, a nebulizer or an atomizer can be used. The formulations, when dispensed, preferably have a particle size in the range of 10 to 100 μ .

[0097] The formulations may conveniently be presented in dosage unit form and may be prepared by any of the methods well known in the art of pharmacy. By the term "dosage unit" is meant a unitary, i.e. a single dose which is capable of being administered to a patient as a physically and chemically stable unit dose comprising either the active ingredient as such or a mixture of it with solid or liquid pharmaceutical diluents or carriers.

2-Alkylidene-18,19-Dinor Slow Release Compounds

[0098] Modified vitamin D compounds that exhibit a desirable and highly advantageous pattern of biological activity in vivo, namely, the more gradual onset and more prolonged duration of activity, may also be used herein.

[0099] Structurally, the key feature of the modified vitamin D compounds having these desirable biological attributes is that they are derivatives of 2-alkylidene-18,19-dinor-vitamin D analogs, in which a hydrolyzable group is attached to the hydroxy group at carbon 25 and, optionally, to any other of the hydroxy groups present in the molecule. Depending on various structural factors -- e.g. the type, size, structural complexity -- of the attached group, these derivatives hydrolyze to the active 2-alkylidene-18,19-dinor-vitamin D analog, at different rates in vivo, thus providing for the "slow release" of the biologically active vitamin D compound in the body.

[00100] The "slow release" in vivo activity profiles of such compounds can, of course, be further modulated by the use of mixtures of derivatives or the use of mixtures consisting of one or more vitamin D derivative together with underivatized vitamin D compounds.

[00101] It is important to stress that the critical structural feature of the vitamin derivatives identified above is the presence of a hydrolyzable group attached to the hydroxy group at carbon 25 of the molecule. The presence of a hydrolyzable group at that position imparts on the resulting derivatives the desirable "slow-release" biological activity profile mentioned above. Other hydroxy functions occurring in the molecule (e.g. hydroxy functions at carbons 1 or 3) may be present as free hydroxy groups, or one or more of them may also be derivatised with a hydrolyzable group.

[00102] The "hydrolyzable group" present in the above-mentioned derivatives is preferably an acyl group, i.e. a group of the type Q^1CO- , where Q^1 represents hydrogen or a hydrocarbon radical of from 1 to 18 carbons that may be straight chain, cyclic,

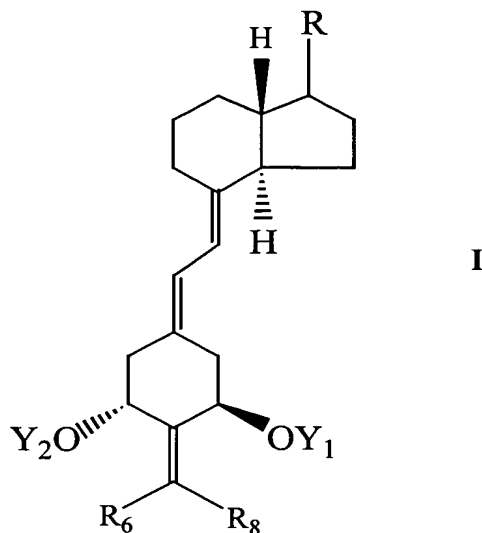
branched, saturated or unsaturated. Thus, for example, the hydrocarbon radical may be a straight chain or branched alkyl group, or a straight chain or branched alkenoyl group with one or more double bonds, or it may be an optionally substituted cycloalkyl or cycloalkenyl group, or an aromatic group, such as substituted or unsubstituted phenyl, benzyl or naphthyl. Especially preferred acyl groups are alkanoyl or alkenoyl groups, of which some typical examples are formyl, acetyl, propanoyl, hexanoyl, isobutyryl, 2-butenoyl, palmitoyl or oleoyl. Another suitable type of hydrolyzable group is the hydrocarbyloxycarbonyl group, i.e. a group of the type $Q^2-O-CO-$, where Q^2 is a C_1 to C_{18} hydrocarbon radical as defined above. Exemplary of such hydrocarbon radicals are methyl, ethyl, propyl, and higher straight chain or branched alkyl and alkenoyl radicals, as well as aromatic hydrocarbon radicals such as phenyl or benzoyl.

[00103] These modified vitamin D compounds are hydrolyzable in vivo to the active analog over a period of time following administration, and as a consequence regulate the in vivo availability of the active analog, thereby also modulating their activity profile in vivo. The term “activity profile” refers to the biological response over time of vitamin D compounds. Individual modified compounds, or mixtures of such compounds, can be administered to “fine tune” a desired time course of response.

[00104] As used herein the term “modified vitamin D compound” encompasses any vitamin D compound in which one or more of the hydroxy functions present in such a compound are modified by derivatization with a hydrolyzable group. A “hydrolyzable group” is a hydroxy-modifying group that can be hydrolyzed in vivo, so as to regenerate the free hydroxy functions.

[00105] In the context of this disclosure, the term hydrolyzable group preferably includes acyl and hydrocarbyloxycarbonyl groups, i.e. groups of the type Q^1CO- and $Q^2-O-CO-$, respectively, where Q^1 and Q^2 have the meaning defining earlier.

[00106] Structurally, the modified vitamin D compounds encompassed may be represented by the formula I shown below:



where Y_1 , Y_2 , and R are as previously defined herein with respect to formula I with the exception that R^5 in the side chain is $-OY_3$ and Y_3 is an acyl group or a hydrocarbyloxycarbonyl group, as previously defined herein.

[00107] Some specific examples of such modified vitamin D compounds include 2-methylene-18,19-dinor derivatives such as:

[00108] 2-methylene-18,19-dinor- $1\alpha,25(OH)_2-D_3-1,3,25$ -Triacetate where $Y_1=Y_2=Y_3$ and is CH_3CO ;

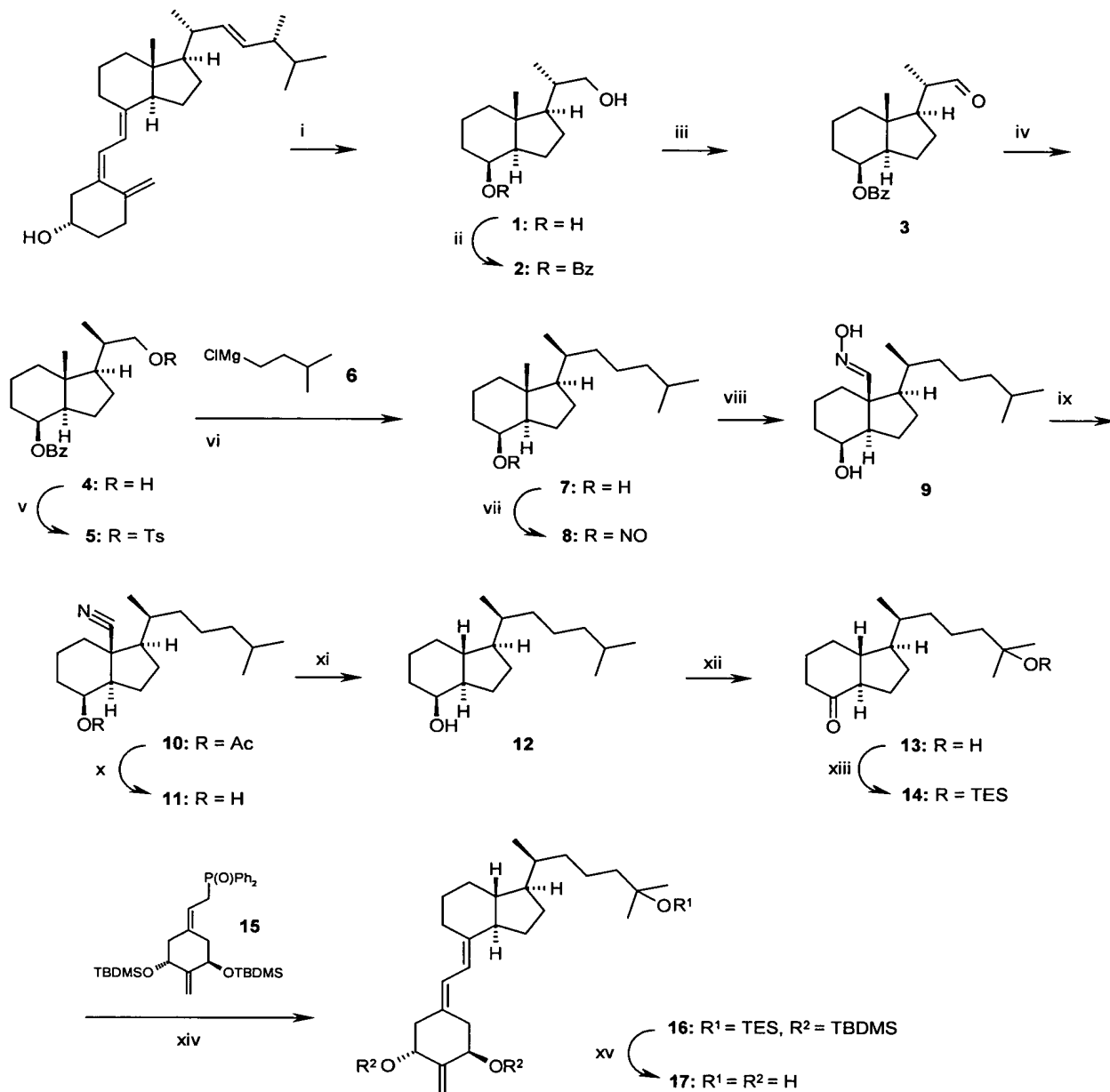
[00109] 2-methylene-18,19-dinor- $1\alpha,25(OH)_2-D_3-1,3,25$ -Trihexanoate where $Y_1=Y_2=Y_3$ and is $CH_3(CH_2)_4CO$;

[00110] 2-methylene-18,19-dinor- $1\alpha,25(OH)_2-D_3-1,3,25$ -Trinonanoate where $Y_1=Y_2=Y_3$ and is $CH_3(CH_2)_7CO$;

[00111] 2-methylene-18,19-dinor- $1\alpha,25(OH)_2-D_3-25$ -Acetate where $Y_1=Y_2$ and is H and Y_3 is CH_3CO .

[00112] These compounds can be prepared by known methods. See for example U.S. Patent 5,843,927.

SCHEME 1



BIOLOGICAL ACTIVITY OF 2-ALKYLIDENE- 18,19-DINOR-VITAMIN D COMPOUNDS

[00113] Figure 1 – Competitive VDR Binding

[00114] Competitive binding of the analogs to the porcine intestinal receptor was carried out by the method described by Dame et al (Biochemistry 25, 4523-4534, 1986).

[00115] TEST MATERIAL

[00116] Protein Source

[00117] Full-length recombinant rat receptor was expressed in E. coli BL21 (DE3) Codon Plus RIL cells and purified using two different column chromatography systems. The first system was a nickel affinity resin that utilized the C-terminal histidine tag on this protein. The protein that was eluted from this resin was further purified using ion exchange chromatography (S-Sepharose Fast Flow). Aliquots of the purified protein were quick frozen in liquid nitrogen and stored at -80°C until use. For use, the protein was diluted in TEDK50 (50 mM Tris, 1.5 mM EDTA, pH7.4, 5 mM DTT, 150 mM KCl) with 0.1 % Chaps detergent so that no more than 20% of the added radiolabeled ligand was bound to the receptor.

[00118] Study Drugs

[00119] Unlabeled ligands were dissolved in ethanol and the concentrations determined using UV spectrophotometry. Serial dilutions were prepared so that a range of unlabeled ligands could be added to the protein without changing the final concentration of ethanol (< 10%) present in the assay mixture. Radiolabeled ligand (3H-1,25(OH)₂D₃) was added in ethanol at a final concentration of 1 nM.

[00120] Assay Conditions

[00121] Radiolabeled and unlabeled ligands were added to 100 µl of the diluted protein, mixed and incubated overnight on ice to reach binding equilibrium. The following day, 100 µl of hydroxylapatite slurry (50%) was added to each tube and mixed at 10-minute intervals for 30 minutes. The hydroxylapatite was pelleted by centrifugation and then washed three times with Tris-EDTA buffer (50 mM Tris, 1.5 mM EDTA, pH 7.4) containing 0.5% Titron X-100. After the final wash,

the pellets were transferred to scintillation vials containing 4 ml of Biosafe II scintillation cocktail, mixed and placed in a scintillation counter. Total binding was determined from the tubes containing only radiolabeled ligand. The percentage of competition was calculated by subtracting the number of dpm remaining in the hydroxylapatite pellet from the total number of dpm bound, dividing by the total number of dpm bound and multiplying by one hundred. Duplicate tubes were prepared and analyzed for each test concentration.

[00122] Figure 2 – HL-60 Cell Differentiation

[00123] The differentiation of HL-60 promyelocytic into monocytes was determined as described by Ostrem et al (J. Biol. Chem. 262, 14164-14171, 1987).

[00124] TEST MATERIAL

[00125] Study Drugs

[00126] The study drugs were dissolved in ethanol and the concentrations determined using UV spectrophotometry (2MD: molar extinction coefficient = 42,000 and λ_{max} = 252nm; 1,25(OH)₂D₃: molar extinction = 18,200 and λ_{max} = 265nm). Serial dilutions were prepared so that a range of drug concentrations could be tested without changing the final concentration of ethanol (< 0.2%) present in the cell cultures.

[00127] Cells

[00128] Human promyelocytic leukemia (HL60) cells were grown in RPMI-1640 medium containing 10% fetal bovine serum. The cells were incubated at 37°C in the presence of 5% CO₂.

[00129] Assay Conditions

[00130] HL60 cells were plated at 1.2 x 10⁵ cells/ml. Eighteen hours after plating, the cells were administered the drug in ethanol. Four days post-dose, the cells were harvested and a nitro blue tetrazolium reduction assay was performed (Collins et al., 1979; J. Exp. Med. 149:969-974, Appendix A). The percentage of differentiated cells was determined by counting a total of 200 cells and recording the number that contained intracellular black-blue formazan deposits. Verification

of differentiation to monocytic cells was determined by measuring phagocytic activity (data not shown). All drug concentrations were tested in duplicate.

[00131] Figure 3 – Transcription Activation

[00132] Transcriptional activity was measured in ROS 17/2.8 (bone) cells that were stably transfected with a 24-hydroxylase (24OHase) gene promoter upstream of a luciferase reporter gene (Arbour et al., 1998). Cells were given a range of doses. Sixteen hours after dosing the cells were harvested and luciferase activities were measured using a luminometer.

[00133] "RLU" in Figure 3 refers to relative luciferase units.

[00134] Figure 4 – Intestinal Calcium Transport

[00135] Weanling, male Sprague-Dawley rats were purchased from Harlan. Upon receipt, the animals were identified by individual tail marks and fed a calcium containing (0.47%) diet (Suda et al., Purified Rodent Diet-Diet 11; Appendix A) for one week before switching to the same diet devoid of calcium (0.02%). Water and a purified rodent diet (Diet 11; Appendix A) containing either 0.47% or 0.02% calcium and 0.3% phosphorus were provided ad libitum. Animals were fed the purified diet containing 0.47% calcium for the first week and then the 0.02% calcium containing diet for the next three weeks of the study. The rats were then fed 0.47% calcium containing diet for one week before switching back to 0.02% calcium containing diet for the remainder of the study. During the second week back on 0.02% calcium containing diet, dose administration began. All doses were administered intraperitoneally in 100 microliters of propylene glycol. Four consecutive doses were given approximately 24 hours apart. Twenty-four hours after the last dose, blood was collected from the tail artery of each experimental animal. The blood was allowed to coagulate at room temperature and then centrifuged at 3000 x g for 15 minutes. The serum was transferred to a polypropylene tube and stored frozen at -20°C. The level of calcium was determined by diluting the serum into 0.1 % lanthum chloride and measuring the absorbance on an atomic absorption spectrophotometer (Perkin Elmer Model 3110,

Shelton, CT). Twenty-four hours after the last dose, intestinal calcium transport was assessed ex vivo using the everted gut sac technique.

[00136] Figures 5 and 6 – Bone Calcium Mobilization

[00137] Weanling, male Sprague-Dawley rats were purchased from Harlan. Upon receipt, the animals were identified by individual tail marks and fed a calcium containing (0.47%) diet (Suda et al., Purified Rodent Diet-Diet 11; Appendix A) for one week before switching to the same diet devoid of calcium (0.02%). Water and a purified rodent diet (Diet 11; Appendix A) containing either 0.47% or 0.02% calcium and 0.3% phosphorus were provided ad libitum. Animals were fed the purified diet containing 0.47% calcium for the first week and then the 0.02% calcium containing diet for the next three weeks of the study. The rats were then fed 0.47% calcium containing diet for one week before switching back to 0.02% calcium containing diet for the remainder of the study. During the second week back on 0.02% calcium containing diet, the animals were tail-bled (baseline serum calcium) and then dose administration was initiated. All doses were administered intraperitoneally in 100 microliters of propylene glycol. Four consecutive doses were given approximately 24 hours apart. Twenty-four hours after the last dose, blood was collected from the tail artery of each experimental animal. The blood was allowed to coagulate at room temperature and then centrifuged at 3000 x g for 15 minutes. The serum was transferred to a polypropylene tube and stored frozen at -20°C. The level of calcium was determined by diluting the serum into 0.1% lanthum chloride and measuring the absorbance on an atomic absorption spectrophotometer (Perkin Elmer Model 3110, Shelton, CT).

INTERPRETATION OF BIOLOGICAL DATA

[00138] Figure 1 illustrates the relative activity of (20S)-2-methylene-18,19-dinor-1 α ,25-dihydroxyvitamin D₃ (also herein referred to as "DP035"), (20S)-2-methylene-19-nor-1 α ,25-dihydroxyvitamin D₃, (also herein referred to as "2MD")

and 1 α ,25-dihydroxyvitamin D₃ (also herein referred to as "C001") in binding to the 1 α ,25-dihydroxyvitamin D pig intestinal nuclear receptor. Figure 1 shows that (20S)-2-methylene-18,19-dinor-1 α ,25-dihydroxyvitamin D₃ is very active in binding to the 1 α ,25-dihydroxyvitamin D₃ receptor from porcine intestinal nuclei.

[00139] The 2-alkylidene-18,19-dinor compounds of this invention exhibit a pattern of biological activity having high potency in promoting the differentiation of malignant cells, relatively high intestinal calcium transport activity and a relatively low ability to mobilize calcium from bone. This is illustrated by the biological assay results obtained for (20S)-2-methylene-18,19-dinor-1 α ,25-dihydroxy-vitamin D₃ which is summarized in Figures 2 through 6. Figure 2 shows a comparison of the activity of the known active metabolite 1 α ,25-dihydroxyvitamin D₃ (C001) as well as analog 2MD and the presently claimed (20S)-2-methylene-18,19-dinor-1 α ,25-dihydroxyvitamin D₃ (DP035) in inducing the differentiation of human leukemia cells (HL-60 cells) in culture to monocytes. Differentiation activity was assessed by a standard differentiation assay, abbreviated as NBT reduction (nitroblue tetrazolium reduction). The assay was conducted according to known procedures, as given, for example, by DeLuca et al U.S. Patent No. 4,717,721 and Ostrem et al, J. Biol. Chem. 262, 14164, 1987. For the assay, the differentiation activity of the test compounds is expressed in terms of the percent of HL-60 cells having differentiated to normal cells in response to a given concentration of test compound.

[00140] The results summarized in Figure 2 clearly show that the analog, (20S)-2-methylene-1 α ,25-dihydroxy-18,19-dinor-vitamin D₃ (DP035) is more potent than 1 α ,25-dihydroxyvitamin D₃ (C001) in promoting the differentiation of leukemia cells. Thus, in the NBT assay close to 90% of the cells are induced to differentiate by 1 α ,25-dihydroxyvitamin D₃ (C001) at a concentration of 1x10⁻⁷M, and the same degree of differentiation is achieved by the (20S)-2-methylene-18,19-dinor analog (DP035) at 1x10⁻⁷M.

[00141] Figure 3 illustrates that (20S)-2-methylene-18,19-dinor-1 α ,25-dihydroxyvitamin D₃ (DP035) has higher transcriptional activity than 1 α ,25-dihydroxyvitamin D₃ in bone cells. This result, together with the cell differentiation activity of Figure 2, suggests that DP035 will be very effective in psoriasis because it has direct cellular activity in causing cell differentiation and in suppressing cell growth. These data also indicate that DP035 may have significant activity as an anti-cancer agent, especially against leukemia, colon cancer, breast cancer, skin cancer and prostate cancer.

[00142] Figures 4 through 6 show a comparison of the calcemic activity of the known active metabolite 1 α ,25-dihydroxyvitamin D₃ (C001), and the 19-nor analog 2MD and the presently claimed (20S)-2-methylene-18,19-dinor-1 α ,25-dihydroxyvitamin D₃ (DP035). Figure 4 shows that (20S)-2-methylene-18,19-dinor-1 α ,25-dihydroxyvitamin D₃ (DP035) is as active as 1 α ,25-dihydroxyvitamin D₃ (C001) in intestinal calcium transport activity. Also, Figures 5 and 6 show that although (20S)-2-methylene-18,19-dinor-1 α ,25-dihydroxyvitamin D₃ (DP035) has some ability to mobilize calcium from bone, it is clearly not as active in this regard as 1 α ,25-dihydroxyvitamin D₃ (C001). Thus, in summary, the (20S)-2-methylene-18,19-dinor analog (DP035) shows a selective activity profile combining high potency in inducing the differentiation of malignant cells, relatively high intestinal calcium transport activity and relatively low bone calcium mobilization activity.